NEUROCHEMICAL CHARACTERISATION OF SENSORY AND AUTONOMIC ENDINGS IN THE MOUSE BLADDER



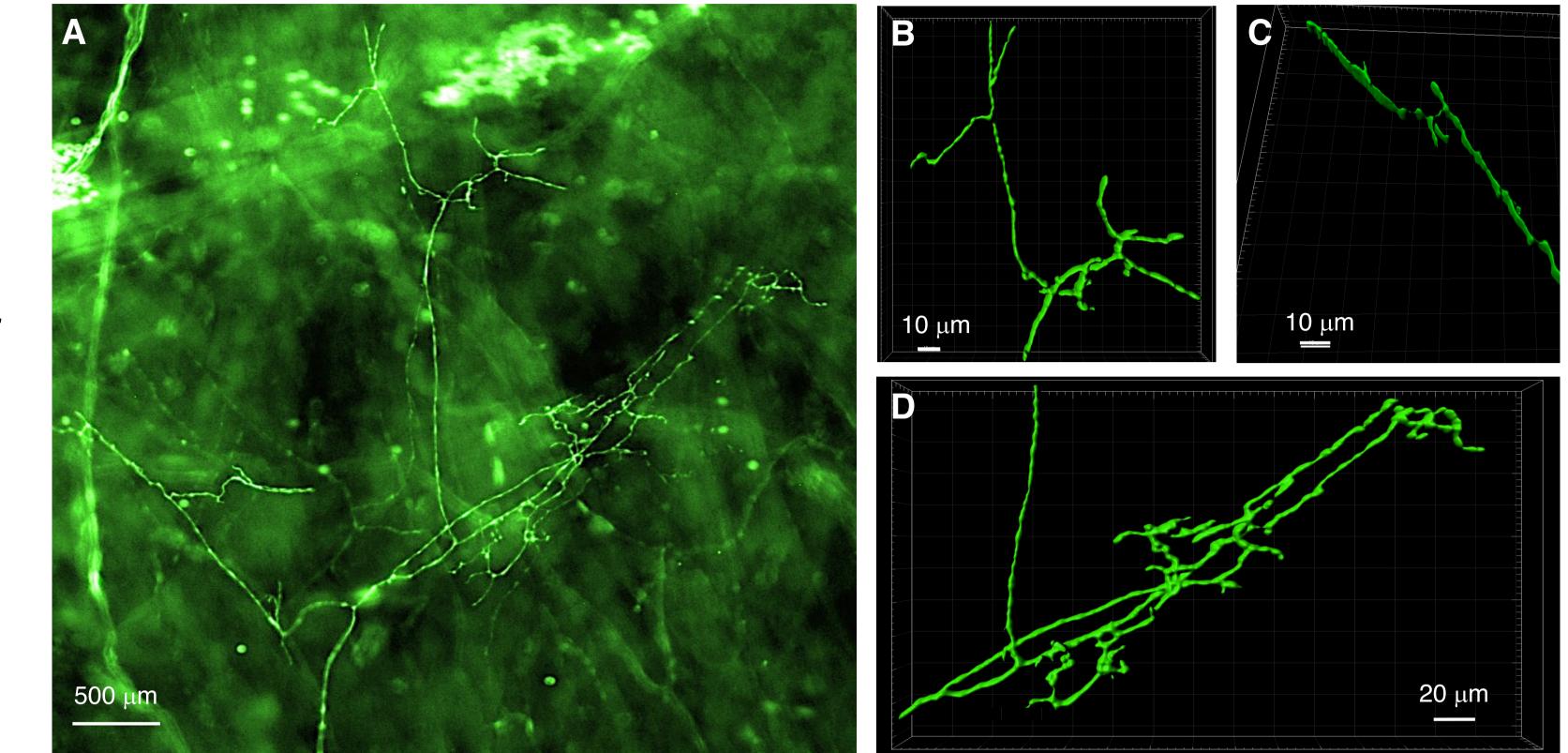
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INTRODUCTION

The bladder is innervated by spinal afferent neurons with cell bodies in the lumbosacral and thoracolumbar dorsal root ganglia (DRG) which project to the bladder via splanchnic-pelvic and lumbarhypogastric nerves. However, there is still a gap in our understanding with regards to comprehensive classification of the of major functional classes of primary afferent neurons in the bladder.

RESULTS

Anterograde labelling (ex vivo) and different types of endings in the muscle layer of mouse bladder



AIMS

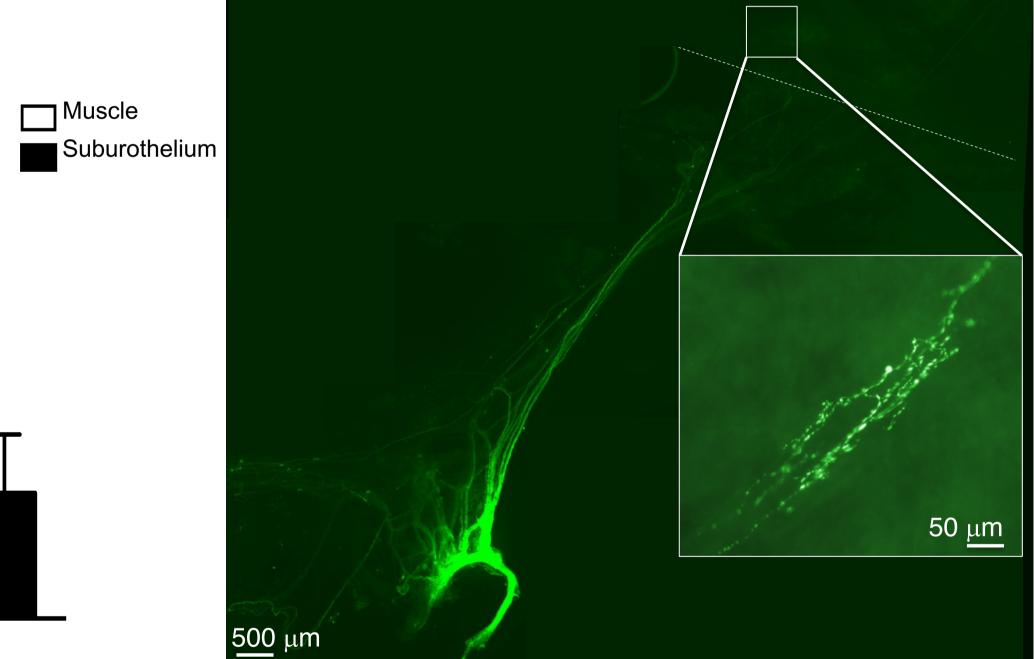
The objective of this study was to characterize anatomically and terminal immunohistochemically the endings of sensory and autonomic neurons in the wholemounts of the mouse bladder.

A- Fluorescence microscopic image at low magnificication, showing the biotinamide labelled parent axon and the associated different types of endings in the muscle layer of the bladder. 3-D reconstruction of z-stacks using laser scanning confocal microscopy of B- simple type ending Ccomplex type ending and D- branching type ending as shown in A.

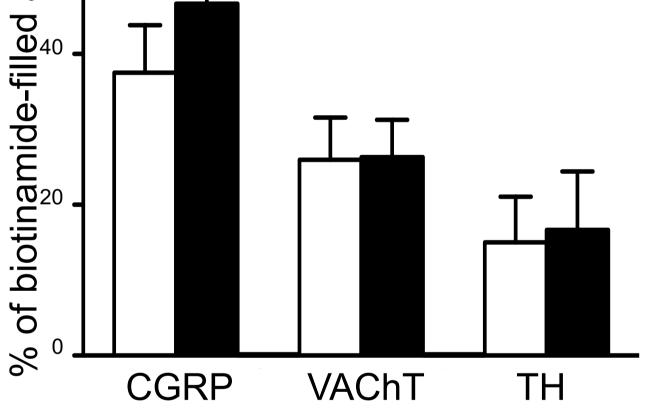
Analysis of co-localisation of biotinamide labelled fibres with different markers

METHODS

We combined anterograde labelling of SUOX pelvic and hypogastric nerves ex vivo and anterograde labelling from lumbosacral



dorsal root ganglia (DRG) in vivo with immunohistochemistry for major markers sensory (calcitonin gene-related Of peptide, CGRP), sympathetic (tyrosine hydroxylase, TH) and parasympathetic (vesicular acetylcholine trasporter, VAChT) nerves. In ex vivo anterograde tracing experiments (Zagorodnyuk & Brookes, 2000), biotinamide was applied to to nerve trunks of the vesical plexus. In in vivo experiments, dextran biotin was injected in L5-S2 DRG of C57BL/6 mice, and a week later was visualised with streptavidin CY3 (Spencer et al, 2018). For triple labelling immunohistochemistry, neurofilament 200 (NF200), CGRP and either vesicular glutamate transporter 2 (VGLUT2) or substance P (SP) was



biotinamide-filled axons. n = 3-6 animals

Group data of biotinamide-filled nerve fibres in Montage of biotinamide labelled hypogastric the muscle and suburothelium layers nerve, projecting into the muscle layer of the combined with three immunohistochemical bladder, edge of the bladder shown by the white markers to identify different populations. dashed line. Expanded region in the white box Values are expressed as percentage of shows a branching type ending in the muscle layer.

CONCLUSIONS

- Immunohistochemical labelling was combined with anterograde labelling ex vivo and in vivo to identify extrinsic nerve endings in the bladder.
- This approach distinguished spinal afferent and autonomic efferent (motor) nerve endings in the bladder.
- Morphology alone (without immunohistochemistry), could not reliably distinguish simple and branching endings of spinal afferent and autonomic

applied to fixed bladder tissue samples.

efferent neurons.

REFERENCES

Sharma H, Kyloh M, Brookes SJH, Costa M, Spencer NJ, Zagorodnyuk VP (2020) Morphological and neurochemical characterisation of anterogradely labelled spinal sensory and autonomic nerve endings in the mouse bladder. J Aut Neu 227: 102697.

Spencer N, Greenhaigh S, Kyloh M, Hibberd TJ, Sharma H, Grundy L, Brierley S, Harrington A, Beckett E, Brookes S, Zagorodnyuk V (2018). Identifying unique subtypes of spinal afferent nerve endings within the urinary bladder of mice. J Comp Neurol 526: 707-720.

Zagorodnyuk V & Brookes S (2000). Transduction sites of vagal mechanoreceptors in the guinea pig esophagus. J Neurosci 20: 6249-55.





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